

Evaluation of the Bioactivities of Plant Extractions from *Lavandula stoechas* L. and *Phlomis syriaca* Boiss on Some Pathogenic Microorganisms Isolated from Local Fresh Red Meat

تقييم الفعالية البيولوجية للمستخلصات النباتية لنباتي (الصَّرمُ المکور) *Phlomis syriaca* و(الأذينة) السورية *Lavandula stoechas* على بعض الكائنات الدقيقة الممرضة المعزولة من اللحوم الحمراء المحلية

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ABSTRACT

Plant extracts have become increasingly important additives in the food industry because of their anti-microbial capacity in processed meat products due to their natural origin. They are an excellent candidate to replace synthetic molecules, which are generally considered to have toxic and carcinogenic effects. The effective extraction of these antioxidant molecules from their natural sources and identifying their activity in commercial products is a major challenge for researchers and contributors to food processing. Pathogenic bacteria were isolated from red meat and study the effect of water extracts of *L. stoechas* and *Phlomis syriaca* plants in different concentrations on pathogens. The application of plant extracts to improve shelf life and nutritional and health characteristics of red meat products. The activities of the water extracts of the *L. stoechas* wild plant on the bacteria isolated from meat. The water extract of the *L. stoechas* plant, with a concentration of 1%, showed positive results on all bacteria ranging between 11 and 15 mm. The sensitivity to the *Klebsiella* extract seemed higher than other bacterial strains, where the inhibition halo was 15 mm in diameter. In contrast, the *Listeria* was resistant to the *Lavandula* water extract with concentrations of 0.5 and 0.25%. The 1% concentration of the water extract of the *Phlomis syriaca* also showed positive results on all bacteria ranging from 10 to 15 mm. The sensitivity of the *Pseudomonas aeruginosa*, *shigella* and *salmonella* appeared high in terms of the extract compared to other bacterial strains. The halo of inhibition was 15 mm with a concentration of 1% of the water extract. For the *Phlomis syriaca*, *Listeria*, *Escherichia coli*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* were resistant to the water extract of the *Phlomis syriaca* at a concentration of 0.25%.

Keywords: Wild plants, *Lavandula stoechas*, *Phlomis syriaca*, red meat, water extracts, pathogenic microbes.

المخلص

تستأثر المستخلصات النباتية بأهمية متزايدة كمضافات مهمة في صناعة المواد الغذائية؛ بسبب مقدرتها المضادة للجراثيم في منتجات اللحوم الجاهزة للأكل، فهي مرشحة ممتازة لتحل محل

الجزيئات المُصنَّعة ذات التأثير السام والمسرطن، وكان الاستخراج الفعال لهذه الجزيئات المضادة للأوكسدة من مصادرها الطبيعية إلى جانب تحديد نشاطها في المنتجات التجارية يمثل تحدياً كبيراً للباحثين والمساهمين في التصنيع الغذائي. تم عزل الأحياء الدقيقة الممرضة من اللحوم ودراسة تأثير المستخلصات المائية لنباتي الضرم المكور والأذينة السورية بتراكيز مختلفة على الأحياء الدقيقة الممرضة. نشاطات المستخلصات المائية لنبات (الضرم المكور *L.stoechas*) البرية على الجراثيم المعزولة من اللحوم الحمراء، حيث أبدى المستخلص المائي لنبات (الضرم المكور) بتركيز (1%) نتائج إيجابية على الجراثيم جميعها تراوحت بين (11 و 15 مم) وبدت حساسية (*الكلبسيلا Klebsiella*) عالية للمستخلص مقارنة بالسلالات الجرثومية الأخرى حيث بلغ قطر هالة التثبيط (15 مم)، في حين أبدت *Listeria* مقاومة للمستخلص المائي للضرم بالتركيزين (0.5 و 0.25%). كما أظهر التركيز (1%) للمستخلص المائي (للأذينة) السورية نتائج إيجابية على الجراثيم تراوحت بين (10 و 15 مم)، وبدت حساسية الزائفة الزنجارية والشيغلة والسلمونيلة عالية للمستخلص مقارنة بالسلالات الجرثومية الأخرى، وبلغ قطر هالة التثبيط (15 مم) بتركيز (1%) للمستخلص المائي للأذينة السورية، وأبدت *الليستيرية والإشريكية القولونية* والزائفة الزنجارية والعنقوديات الذهبية مقاومة للمستخلص المائي (للأذينة) السورية بتركيز (0.25%). تطبيق المستخلصات النباتية لتحسين مدة الصلاحية، والخصائص الغذائية والصحية لمنتجات اللحوم الحمراء.

الكلمات المفتاحية: الفصيلة الشفوية، الضرم المكور، الأذينة السورية، اللحوم الحمراء، المستخلصات المائية، الجراثيم الممرضة.

INTRODUCTION

Food poisoning is one of the most common causes of disease and death in developing countries (Doughari and Pukuma, 2007; Pirbalouti A. G. *et al.*, 2010; Sapkota R. *et al.*, 2012) due to contamination with gram-negative bacteria, such as *Salmonella typhi* and *Escherichia coli*, *Escherichia coli*, *Pseudomonas aeruginosa* (Solomakos N *et al.*, 2008; Pandey A., Singh P. 2011), and Gram-positive bacteria, such as *Staphylococcus aureus* and *Bacillus cereus*.

However, preservative chemicals can be used to prevent bacterial growth and prevent food spoilage (Yamamura A *et al.*, 2000). The control of food-borne pathogens requires the use of various preservation techniques in the manufacture and storage of food products. Therefore, there is an increasing need for active

preservatives that can prolong product life, inhibiting non-salt microbial growth and sugar, which the consumer prefers at low levels (Zink, 1997). The negative perception of synthetic food additives and the emergence of negative side effects in health led to the development of natural alternatives and the use of extracts pates and oils antibacterial agents, which are safer and more active (Bialonska D et al., 2010, Tajkarimi M. M. et al., 2010; Hussain M et al., 2018).

On the other side, essential oils or their components have antibacterial properties (Oussalah M et al., 2007), parasites (George et al., 2009), viruses (Astani A. et al., 2011), fungi (Silva F et al., 2011; Tiemersma et al., 2011) and antifungal agents. However, the use of vegetable oils and extracts as food preservatives requires knowledge of their Minimal Inhibition Concentration (MIC).

Meat products are oxidized, especially when stored. Mincing facilitates the interaction of oxidants with unsaturated fatty acids and the formation of free, which negatively affects color, flavor, texture, and food quality (Vaithiyanathan S et al., 2011). Fortunately, the application of antioxidants prevents fat oxidation (Martinez-Tome et al., 2001), and plants are of great importance as natural additives (Lindberg, Bertelsen 2011; Zheng and Wang, 2001).

RESEARCH PROBLEM

There is a lack of local studies on wild plants, despite their spread in different Syrian environments. It hinders efforts in determining the extent of the effect of their extracts on pathogenic or non-pathogenic bacteria. It also afflicts the utilization of plant compounds as substances with biological effectiveness and as alternatives to biological antibiotics.

RESEARCH SIGNIFICANCE

Natural alternatives are important for preserving food, such as plant extracts and essential oils with their anti-microbial properties that have a negative effect on food composition and their nutritional quality. Therefore, studying the effect of Syrian lavender and atrium as meat preservatives, preventing microorganism corruption or transfer to pathogens, and prolonging their storage is of great importance.

RESEARCH AIMS

The aim of this research is to identify the anti-aqueous activity of *L. stoechas* plants and *Phlomis syriaca* on some bacteria isolated from red meat and preserve them as bioactive and alternative substances for antibiotics and synthetic chemicals.

MATERIALS AND METHODS

Plant Materials

The samples of the *L. stoechas* and *Phlomis syriaca* selected for the research were collected in the village of Qismin from the North Kabir area below Tishreen Lake, west of the riverbed at an altitude of about 100-170 m above sea level, 20 km northeast of Lattakia and north of Aleppo road. The sample was sent to the laboratory, washed with sterile distilled water, and then dried on filter paper at 25°C.

L. stoechas, in Figure 1, belongs to the oral family Oblige letter, a herb that spreads in the Mediterranean and the Black Sea areas, on sunny slopes and favors silicic soils. It does not grow in the shade and is relatively tolerant to drought. It is a small branched shrub of 30-50 cm long and has flowers. In general, the coronets are purple, flowering from February to May (ACSAD, 2012).



Figure 1. *Lavandula stoechas* L.

The Syrian atrial plant *Phlomis syriaca*, Figure 2, belongs to the oral family. This genus' species home is the Mediterranean basin and from Central Asia to China. The flowers are yellow or violet and bloom during the period from April to June. Various studies have indicated the medicinal value of the plant. It enhances the protection of the liver, kidneys, bones, heart, and blood vessels. It is also antibacterial, anti-allergic, and has antioxidant properties (Demirci et al., 2008).



Figure 2. Part of *Phlomis syriaca* Boiss



Figure 3. Lamb and veal samples

Preparation of Plant Extracts

Aqueous extracts

Previously dried leaves were placed in the incubator for two hours at 37°C to eliminate any moisture in the sample for easy grinding of the leaves (Blazekovic et al., 2011). The dry leaves were ground with an electric grinder, 25g of the powder was taken and soaked in a 500ml flask with 250ml of distilled water (Khayyat et al., 2018). The samples were placed in the shaker at 170 rpm at laboratory temperature (25°C), away from light for 48 hours (Mahmoudi et al., 2014). The extract was filtered using a 0.45 Watman filter paper and dried in an air incubator at a temperature not exceeding 40°C.

Isolation of pathogens from red meat

The researchers took 10 samples of lamb and 10 samples of veal, Figure 3. Ten grams of each sample were added to 100 ml of sterile peptone, then mixed for 3 minutes at high speed, from 10-1 to 10-4, after they have compiled the analysis of the Hungarian neighborhoods (i.e., vaccination).

Pathogenic microorganisms were isolated from meat samples and a selection of selective solid bacterial media was taken, such as EMB Agar for *Enterobacter* development, Chapman Agar for staphylococcal development, and Cetramide Agar to characterize *Pseudomonas aeruginosa* Editing ammonia, Agar PDA Dextrose Potato medium for the development of *Candida albicans*, as well as nutritious agar medium and nutrient broth (APHA, 2000). Bacteria isolated from the samples could be classified after conducting the necessary biochemical tests (oxidase, catalase, indole, methyl red, citrate fermentation, VogtsProscaur, H₂S release) based on the Bergi evidence (Garrity G. M et al., 2005).

Furthermore, the sensitivity and resistance of isolated microbes were tested by the propagation method of tablets. The sensitivity and resistance of some antibiotics were determined by measuring halos inhibitors on Mueller Hinton Agar's medium (Barker and Kehoe E, 1995).

A group of antibiotics was used (Table 1) with a concentration in the tablets.

Table 1 Used Antibiotics, mcg.

Antibiotic	Code
Ampicillin/ Cloxacillin	APX: 25/5 mcg
Lincomycin	L:2 mcg
Erythromycin	E:15 mcg
Cefaclor	CEC: 30 mcg
Pipemidic	PI:20 mcg

Preparation of bacterial inoculum

The activation of activated microbes was carried out on the Nutrient Agar NA and incubated at 37°C for 24 hours. Ten colonies of each type of bacterium were transferred, under sterile conditions, to a test tube containing 5 ml of broth nutrients. At 37°C for 4 hours, appropriate dilutions were carried out for each type of bacterium, with a total cell count of 1.5×10^{10} cells/ml (Anandiand Juan, 2009).

Test of anti-microbial Bioactivity of Plant Extraction

Aqueous extracts were prepared at a concentration of 0.25, 0.5, 0.75, and 1.0%. Muller-Hinton medium was prepared by dissolving 38 g of it in a liter of distilled water, with 0.5 ml of bacterial suspension, and brushes were transferred over the culture medium with a cotton swab. After 15 minutes, the extracts were distributed and placed in the fridge. The incidence of inhibition zones is evidence of bacterial growth inhibition, and their diameters are measured after finishing

the lap with a precise measuring instrument or millimeter ruler. The experiment was performed with three replications (Kelmanson et al., 2000).

RESULTS AND DISCUSSION

The Effect of Dry Extracts of *Lavandula stoechas* L. and *Phlomis syriaca*

The results shown in Table 2 showed that the *L. stoechas* plant contains 22.98% of the pure extract of aqueous extract. In comparison, the *Phlomis syriaca* plant contains 21.11% of the pure extract.

Table 2 The Yield of Dry Plant Extracts Used in the Research.

Plant	the sample	Repeater%			Average and deviation
		1	2	3	
<i>Lavandula stoechas</i>	25	23.56	22.4	22.98	22.98 ± 0.58
<i>Phlomis syriaca</i>	25	20.5	21.73	21.11	21.11 ± 0.61

Bacterial species isolated from meat

Table 3 shows the six isolated bacterial species when testing red meat microbiologically and one type of yeast *Candida albicans*. See the *Pseudomonas aeruginosa*, *E.coli* in Figure 5, the *Klebsiella* in Figure 4 and the *Shigella* (Table 3).

Table 3 Microorganisms Isolated from Red Meat from Different Areas in Homs

Microbe strain	Rate of positive samples in meat%	
	Sheep meat	veal
<i>E. coli</i>	85.71	66.66
<i>Pseudomonas aeruginosa</i>	57.14	33.33
<i>Klebsiella sp.</i>	42.85	16.66
<i>Shigella sp.</i>	28.57	50.00
<i>Salmonella sp.</i>	42.85	33.33
<i>Staphylococcus aureus</i>	14.28	66.66
<i>Candida albicans</i>	28.57	16.66

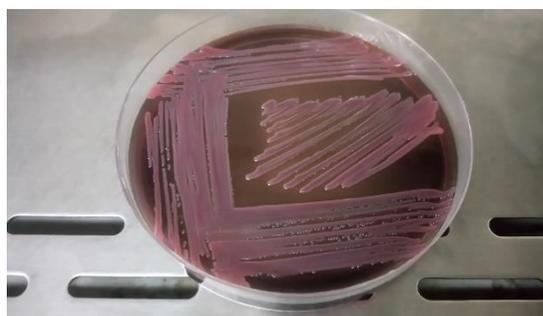


Figure 4. *Klebsiella* on EMB

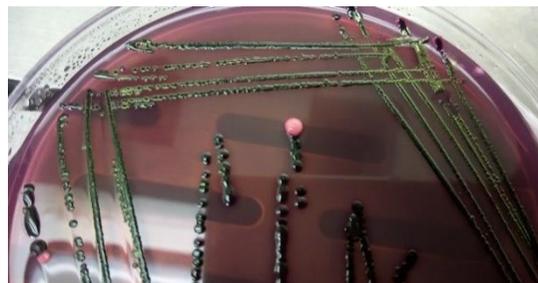


Figure 5. *E. coli* on EMB

Biological activity of *L.stoechas* plant extracts on pathogenic microorganisms.

Table 4 shows the results of the study of the biological activity of water extract of the *L. stoechas* against germ

Table 4 Diameter of Halo Inhibition of Aqueous Extract of the Wild Plant *L. stoechas* for Tested Germs and *Candida albicans*, per mm

Microbe strain	Average halo inhibition diameter,%			
	0.25	0.5	0.75	1.0
<i>E. coli</i>	0.0 ± 7	9 ± 0.0	12 ± 1.0	14 ± 0.57
<i>Pseudomonas aeruginosa</i>	6 ± 0.0	10 ± 1.0	11 ± 0.57	13 ± 1.0
<i>Klebsiella sp.</i>	-	8 ± 1.52	12 ± 0.0	15 ± 1.52
<i>Shigella sp.</i>	-	09 ± 1.0	11 ± 0.57	12 ± 2.08
<i>Salmonella sp.</i>	6 ± 1.0	8 ± 0.57	12 ± 0.57	14 ± 1.52
<i>Staphylococcus aureus</i>	7 ± 1.0	9 ± 0.0	12 ± 1.0	13 ± 0.0
<i>Listeria sp.</i>	-	-	9 ± 1.0	11 ± 0.0
<i>Candida albicans</i>	7 ± 0.57	10 ± 1.0	11 ± 0.0	14 ± 0.0

Moreover, the concentration of 1% showed positive results on all bacteria ranging from 11 to 15 mm (Figure 6). The *klebsiella* sensitivity of the extract was high compared to other bacterial strains. The inhibition of 9 mm in concentration was 0.75%, which is consistent with the result of the researchers (Tabatabaei et al., 2014). The maximum value (14 mm) was 1.0%.



Figure 6. The efficacy of *L. stoechas* water extract on *E.coli*

Table 5 shows the results of the study of the impact of the biological activity of the aqueous extract of the *Boiss* against bacteria.

Table 5 Diameter of Halo Inhibition of Aqueous Extract of Syrian Atrial Plant *Phlomis syriaca* for Tested Bacteria and *Candida albicans*, per mm

Microbe strain	Average halo inhibition diameter,%			
	0.25	0.5	0.75	1
<i>E. coli</i>	-	7± 0.0	9± 1.0	10± 0.0
<i>Pseudomonas aeruginosa</i>	-	8± 0.57	10± 0.0	15± 1.0
<i>Klebsiella sp.</i>	6± 1.0	7± 1.52	9± 0.0	10± 1.52
<i>Shigella sp.</i>	-	9± 1.0	11 ± 0.57	15± 2.08
<i>Salmonella sp.</i>	06 ± 1.0	8± 0.0	10± 0.57	15± 1.52
<i>Staphylococcus aureus</i>	-	7± 0.0	10± 1.0	12± 0.0
<i>Listeria sp.</i>	-	6± 1.0	8± 1.0	10± 0.0
<i>Candida albicans</i>	6±1.0	9± 1.0	11 ± 0.0	14 ± 0.0

The concentration of 1% showed positive results on all germs ranging from 10 to 15 mm. The sensitivity of *Pseudomonas aeruginosa* in Figure 7. The *Shigella* and *Salmonella* appeared to be high for the extract compared to other bacterial strains. Gold resistance to aqueous extract of the Boiss was at a concentration of 0.25%.

Table 6 shows the results of the sensitivity of the isolated pathogenic bacteria to the tested biological antibiotics. All the bacteria under test were resistant to Ampicillin, Cefaclor, Lincomycin, and Erythromycin.

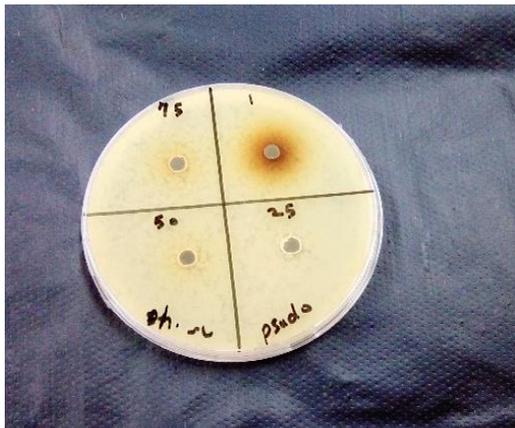


Figure 7. The efficacy of *Phlomis syriaca* water extract on *Pseudomonas aeruginosa*

Table 6 Results of Sensitivity and Resistance of Microorganism Strains to Biological Antibiotics, Inhibition Diameter, 1 mm

abbrev. Biological R code	APX	CEC	L	E	PI
conc.	30	30	2	15	20
<i>E. coli</i>	R	R	R	R	6
<i>Pseudomonas aeruginosa</i>	R	R	R	R	9
<i>Klebsiella</i>	R	R	R	R	9
<i>Shigella</i>	R	R	R	R	9
<i>Salmonella</i>	R	R	R	R	8
<i>Staphylococcus aureus</i>	R	R	R	R	9
<i>Listeria</i>	R	R	R	R	20
<i>Candida albicans</i>	R	R	R	R	9
Resistance = R					

The gram-positive *Listeria* bacteria showed high sensitivity to Tobramycin and Pipemidic, with a damping aura of 16 and 20 mm, respectively. The gram-negative *Pseudomonas*

aeruginosa showed good sensitivity against these inhibitors.

The results show that the aura diameters of aqueous extracts were generally larger than the diaphragms of the biological antibiotics used.

CONCLUSION

1. Aquatic extracts of *L. stoechas* and *Boiss* are anti-microbial agents.
2. The aura diameters of inhibition of aqueous extracts were generally greater than the damping aura diameters of the antibiotics used.
3. Using plant extracts can be a healthy method and technique for preserving meat.

Botanical extracts are considered beneficial nutrients for health because they contain medicinal compounds

REFERENCES

- ACSAD (2012). *Atlas of Medicinal and Aromatic Plants in the Arab World, Damascus, Syria*, 356 - 357.
- Anandi M., Juan C. (2009). Drug susceptibility testing for *Mycobacterium tuberculosis*. *Journal of Antimicrobial Chemotherapy*. 54:130-133.
- APHA, AWWA and WEF (2000). *Standard Methods for Examination of water and wastewater*. 20th edition. American Public Health Association, Inc., Baltimore, M. D. USA.
- Astani A., Reichling J., Schnitzler P. (2011). Screening for antiviral activities of isolated compounds from essential oils. *Evidence-based complementary and alternative medicine*.
- Barker G. A.; Kehoe E. (1995). Assessment of disc diffusion methods for susceptibility testing of *Aeromonas salmonicida*. *Aquaculture*, Vol. 134:1-8.
- Bialonska D., Ramnani P., Kasimsetty S., Muntha K., Gibson G., Ferreira D. (2010). The influence of pomegranate by-product and punicalagins on selected groups of human intestinal microbiota. *Int. J. Food Microbiol.* 140:175-182.
- Blazekovic B., Stanic G., Pepeljnjak S., Vladimir-Knezevic S. (2011). In vitro antibacterial and antifungal activity of *Lavandula × intermedia* Emeric ex Loisel. 'Budrovka'. *Molecules*, 16: 4241-4253.
- Doughari J. H., Pukuma M., D. N. (2007). Antibacterial effects of *Balanites aegyptiaca* L. Drel. and *Moringa oleifera* Lam. on *Salmonella typhi*. *African Journal Biotechnology*. 6(19): 2212-2215.
- Demirci, F., K. Guven, B. Demirci, M. Dadandi, K. Baser, (2008) Antibacterial activity of two *Phlomis* essential oils against food pathogens, *Food Control.*, 19, 1159-1164.
- Garrity G. M.; Brenner D.; Krieg N., Staley J. (2005). *Bergey's Manual of Systematic Bacteriology*. Springer, USA, 2nd Ed., Vol. 2, P. 1-1135.
- George D. R., Smith T., Shiel R., Sparagano O., Guy J. (2009). Mode of action and variability in efficacy of plant essential oils showing toxicity against the poultry red mite, *Dermanyssus gallinae*. *Vet. Parasitol.* 161, 276-282.

- Hussain M., Ali Nizam A., Abou-Isba S., Abou-Younes A., Khaddour W. (2018). A Broad-Spectrum Antibacterial Activity of Lyophilized Crude Extracts of *Bacillus subtilis* against Clinical and Food-Borne Pathogens, *International Research Journal of Pharmacy and Medical Sciences (IRJPMS)*, Volume 1, Issue 3, pp. 15-21, 2018.
- Kelmanson J., Jager A., Standen J. (2000). Zulu medicinal plants with antibacterial activity. *J. Ethnopharmacol.*, 69:241-246.
- Khayyat S., Al-Kattan M., Basudan N. (2018). Phytochemical Screening and Antidermatophytic Activity of Lavender Essential Oil from Saudi Arabia. *International Journal of Pharmacology Volume 14* (6): 802-810.
- Lindberg M. H., Bertelsen G. (1995). Spices as antioxidants. *Trends Food Sci. Technol.*, 6, 271-277.
- Mahmoudi R., Zare P., Hassanzadeh P., Nosratpour S. (2014). Effect of *Teucrium polium* essential oil on the physicochemical and sensory properties of probiotic yoghurt. *J Food Process Pres.* 38:880–888.
- Martinez-Tome M., Jimenez A., Ruggieri S., Frega N., Strabbioli R., Murcia M. (2001). Antioxidant properties of Mediterranean spices compared with common food additives. *J. Food Prot.*, 64,1412-1419.
- Oussalah M., Caillet S., Saucier L., Lacroix M. (2007). Inhibitory effects of selected plant essential oils on the growth of four pathogenic bacteria: *E. coli* O157:H7, *Salmonella typhimurium*, *Staphylococcus aureus* and *Listeria monocytogenes*. *Food Control* 18, 414–420.
- Pandey A., Singh P. (2011). Antibacterial activity of *Syzygium aromaticum* (Clove) with metal ion effect against food borne pathogens. *Asian J. Plant Sci. Res.* 1(2): 69-80.
- Pirbalouti A. G., Jahanbazi P., Enteshari S., Malekpoor F., Hamedi B. (2010). Anti-microbial activity of some Iranian medicinal plants. *Arch. Biol. Sci. Belgrade.* 62(3): 633-642.
- Sapkota R., Dasgupta R., Rawat D. (2012). Antibacterial effects of plants extracts on human microbial pathogens and microbial limit tests. *Int. J. Res Pharm. & Chem.*, 2(4): 926-936.
- Silva F., Ferreira S., Duarte A., Mendonça D., Domingues F. (2011). Antifungal activity of *Coriandrum sativum* essential oil, its mode of action against *Candida* species and potential synergism with amphotericin B. *Phytomedicine* 19, 42-47.
- Solomakos N., Govaris A., Koidis P., Botsoglou N. (2008). The anti-microbial effect of thyme essential oil, nisin and their combination against *Escherichia coli* O157:H7 in minced beef during refrigerated storage. *Meat Science.* 80, 159-166.
- Tajkarimi M. M., Ibrahim S., Cliver D. (2010). Anti-microbial herb and spice compounds in food. *Food Control* 21, 1199-1218.
- Tabatabaei Y., Behbehani B., Mortazavi A. (2014). Investigate the minimum inhibitory concentration (MIC) and the minimum antibacterial concentration (MBC) of *Lavandula stoechas* and extracts *Rosmarinus officinalis*. on the pathogenic bacteria “in vitro No.2 ISSN 2008-4978 Vol.5 (JPS).
- Tiemersma E.W., Bronzwaers S., Lyytikäinen O., Degener J., Schrijnemakers P., Bruinsma N., Monen J., Witte W., Grundman H. (2004). European anti-microbial resistance surveillance system participants: Methicillin-resistant *Staphylococcus aureus* in Europe. *Emerg. Infect. Dis.* 10:1627–1634.
- Vaithyanathan S., Naveena B., Muthukumar M., Girish P., Kondaiah N. (2011). Effect of dipping in pomegranate (*Punicagranatum*) fruit juice phenolic solution on the shelf life of chicken meat under refrigerated storage (4 °C). *Meat Sci.*, 88, 409-414.
- Yamamura A., Murai A., Takamatsu H., Watabe K. (2000). Anti-microbial effect of chemical preservatives on enterohemorrhagic *Escherichia coli* O157:H7. *J. Health Sci.* 46: 204-208.
- Zheng W., Wang S. (2001). Antioxidant activity and phenolic compounds in selected herbs. *J. Agric. Food Chem.*, 49, 5165-5170.
- Zink D. L. (1997). The impact of consumer demands and trends on food processing. *Emerging Infect. Dis.* 3, 467-469